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## MURAYAANTHRAQUINONE, A HYBRID BENZ[a]ANTHRAQUINONE FROM A UV MUTANT OF STREPTOMYCES MURAYAMAENSIS

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Abstract: An X-ray crystallographic analysis revealed that a new metabolite of *Streptomyces murayamaensis*, murayaanthraquinone, contains a 5-azo-16-oxo-dibenzo[b,k]chrysene ring system. This hexacyclic structure has been shown to be a hybrid derived from a benz[a]anthraquinone and a 3-amino-4-hydroxybenzoic acid.

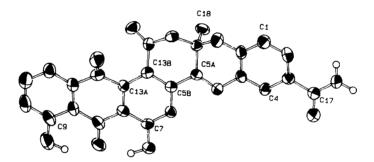
In our continuing studies of kinamycin biosynthesis (e.g. kinamycin D, 1<sup>1</sup>), in *Streptomyces murayamaensis*, <sup>2,3</sup> we have generated a number of UV mutants that accumulate a variety of colored metabolites that are either trace components or are undetectable in extracts of the wild-type organism. Diode array-detected HPLC of EtOAc extracts from the wild-type strain revealed a new metabolite with a UV chromophore (260, 304, 420 nm) similar to ketoanhydrokinamycin, 2.<sup>4</sup> Extracts of blocked strain MC2<sup>5</sup> also contained the new metabolite, but in greater amount. We now report that this new orange-yellow metabolite, 3, is a hybrid composed of a benz[a]anthraquinone and 3-amino-4-hydroxybenzamide, 4.<sup>6</sup>

Two liters of a 7-day fermentation of strain MC2 in 7% farina/0.2% trace metals medium were worked up to the EtOAc extract.<sup>5</sup> Concentration afforded a solid residue (750 mg), which was chromatographed in thirds on phosphate-buffered (pH 7.0) flash grade silica gel (3 x 16 cm, 3% MeOH/CH<sub>2</sub>Cl<sub>2</sub>). Elution with the same solvent yielded a fraction (~12 mg) enriched in 3. This was purified on a column of Sephadex LH-20 (1.8 x 27 cm, 50% MeOH/CH<sub>2</sub>Cl<sub>2</sub>) and afforded a bright orange-yellow fraction containing ~2 mg of pure 3.

High resolution positive ion FABMS furnished the formula C<sub>26</sub>H<sub>17</sub>N<sub>2</sub>O<sub>7</sub> (M + H<sup>+</sup>, m/z 469.1030, calcd 469.1024). Absorptions at 1709, 1669 and 1629 cm<sup>-1</sup> in the IR spectrum indicated a ketone and a quinone (one carbonyl hydrogen-bonded). The <sup>1</sup>H- and <sup>1</sup>H{<sup>1</sup>H}COSY NMR spectra yielded two partial structures, 5 and 6, in addition to a quaternary methyl singlet ( $\delta$ 1.30), a diastereotopic methylene ( $\delta$ 3.15 and 3.90, J = 13 Hz), an aromatic singlet ( $\delta$ 8.10), and a broad exchangeable signal centered at  $\delta$ 11.9. During an attempt to obtain a <sup>13</sup>C NMR spectrum in DMSO  $d_0$ , the sample crystallized. It was re-dissolved by heating, and then slowly cooled over a 48-hour period to furnish larger crystals. X-ray diffraction analysis of a single triclinic needle was performed to a maximum  $\sin\theta/\lambda = 0.5436$  Å<sup>-1</sup>. Using 1336 observed reflections, the space group P-1 (#2), and the direct methods program SHELXS (TEXAN crystallographic software package)<sup>7</sup> the positions of a C<sub>26</sub>N<sub>2</sub>O<sub>7</sub> molecule, along with a molecule of DMSO were determined. Hydrogen atoms were

placed in calculated positions, and a DIFABS<sup>8</sup> absorption correction (transmission factors 0.81 to 1.28) applied. Anisotropic refinement of all non-hydrogen atoms gave R = 0.041 and  $R_w = 0.044$ . Successful refinement in the centrosymmetric space group P-1 demonstrates the crystal contains a racemic mixture of 3.

Figure 1. ORTEP drawing from the single-crystal X-ray structure determination of 3. Hydrogens on carbon have been omitted for clarity.



A  $^{13}$ C-enriched sample 3a, was later prepared biosynthetically by feeding [U- $^{13}$ C6]-D-glucose,  $^{9\cdot11}$  7, (98+% enriched, 1.0 g in 25 mL H<sub>2</sub>O) in thirds at 24, 48, and 72 hours after inoculation, distributed amongst 5 200-mL fermentations. Work-up afforded 1.6 mg of 3a.  $^{13}$ C NMR analysis revealed ~2% enrichments at each of the benzanthraquinone carbons, but extremely low enrichments for those of the aminohydroxybenzamide moiety. Six pairs of  $^{13}$ C- $^{13}$ C coupled resonances were readily matched from the  $^{13}$ CC values and their assignments then made from the chemical shifts. These were confirmed and all but four others assigned (Table 1) from HMQC and HMBC NMR experiments (Scheme 1).

## Scheme 1

HO

$$\begin{array}{c} CHO \\ -OH \\ -OH \\ -OH \\ -CH_2OH \end{array}$$
 $\begin{array}{c} H_3C = COSCOA \end{array}$ 
 $\begin{array}{c} H \\ -OH \\ -OH \\ -OH \\ -OH \\ -OH \end{array}$ 
 $\begin{array}{c} OH \\ OH \\ -OH \\ -OH$ 

| Pos | <sup>1</sup> H NMR                  | <sup>13</sup> C NMR | 1 <sub>JCC</sub> | Pos    | <sup>1</sup> H NMR                 | 13 <sub>C NMR</sub>  | $^{1}J_{CC}$ |
|-----|-------------------------------------|---------------------|------------------|--------|------------------------------------|----------------------|--------------|
| 1   | 7.10, 1H, d, <i>J</i> =8.4 Hz       | 116.3 CH            | *                | 12     | 7.54, 1H, d, <i>J</i> =7.2 Hz      | 118.7 CH             | 61.4         |
| 2   | 7.85, 1H, dd, <i>J</i> =8.4, 2.0 Hz | 131.4 CH            | *                | 12a    |                                    | 135.0 C              | 61.2         |
| 3   |                                     | 129.0 C             | *                | 13     |                                    | 182.6 C              | 53.2         |
| 4   | 8.08, 1H, d, <i>J</i> =2.0 Hz       | 127.0 CH            | *                | 13a    |                                    | ** C                 |              |
| 4a  |                                     | 132.0 C             | *                | 13b    |                                    | ** C                 |              |
| 5a  |                                     | 157.1 C             | 55.7             | 14     |                                    | 192.6 C              | 52.7         |
| 5b  |                                     | ** C                |                  | 15     | 3.15, 3.90, 2H, d, <i>J</i> =13 Hz | 51.5 CH <sub>2</sub> |              |
| 6   | 8.1, 1H, s                          | 117.5 CH            | 68.6             | 15a    |                                    | 74.3 C               | 37.5         |
| 7   |                                     | 162.5 C             | 70.2             | 16a    |                                    | 148.0 C              | *            |
| 7a  |                                     | 121.1 C             | 55.5             | 17     |                                    | 166.7 C              | *            |
| 8   |                                     | 190.0 C             | 55.7             | 18     | 1.3, 3H, s                         | 20.7 CH <sub>3</sub> | 37.5         |
| 8a  |                                     | 115.9 C             | 65.0             | 17-NH2 | 8.00, 2H, br, exchangeable         |                      |              |
| 9   |                                     | 160.7 C             | 64.6             | 7-OH   | 11-12, br, exchangeable            |                      |              |
| 10  | 7.37, 1H, d, <i>J</i> =8.6 Hz       | 124.0 CH            | 57.0             | 9-OH   | 11-12, br, exchangeable            |                      |              |
| 11  | 7.81, 1H, dd, J=8.2, 8.2 Hz         | 137.6 CH            | 56.9             | 1      |                                    |                      |              |

Table 1. <sup>1</sup>H- and <sup>13</sup>C NMR spectroscopic data for 3 and 3a.

- \* The enrichment was very low and JCC could not be measured.
- \*\* 13C chemical shift could not be identified by the HMBC experiment.

The formation of compound 3 can be rationalized by addition of an o-aminophenol to a p-quinone 8 (Scheme 2). The latter is presumably derived from dehydrorabelomycin, 9, previously identified as an S. murayamaensis metabolite. <sup>12</sup> Indeed, 8 (PD 116744) and 9 have been isolated from Streptomyces sp. WP 3668. <sup>13,14</sup> The coupling partner (either the benzoic acid 10 or the benzamide  $4^6$ ) represents a new naturally-occurring regioisomer of aminohydroxybenzoic acid. The 2,3-, <sup>15</sup> 2,6-, <sup>16</sup> and 3,5-isomers <sup>17,18</sup> have all been shown to be derived from the shikimic acid pathway. The origin of 4 will be reported in due course.

## Scheme 2

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